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Nutrition

Adequate energy-protein intake is not enough to improve nutritional and metabolic status in muscle-depleted patients with chronic heart failure

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Abstract

Background:

An adequate energy-protein intake (EPI) when combined with amino acid supplementation may have a positive impact nutritional and metabolic status in patients with chronic heart failure (CHF).

Methods and results:

Thirty eight stable CHF patients (27 males, 73.5 ± 4 years; BMI 22.5 ± 1.4 kg/m²), with severe depletion of muscle mass and were randomised to oral supplements of essential amino acids 8 g/day (EAA group; $n=21$) or no supplements (controls; $n=17$). patients had adequate EPI (energy ≥ 30 kcal/kg; proteins >1.1 g/kg). At baseline and 2-months after randomisation, the patients underwent metabolic (plasma lactate, pyruvate concentration; serum insulin level; estimate of insulin

resistance by HOMA index), nutritional (measure of nitrogen balance), and functional (exercise test, walking test) evaluations.

Body weight increased by >1 kg in 80% of supplemented patients (mean 2.96 kg) and in 30% of controls (mean 2.3 kg) (interaction <0.05). Changes in arm muscle area, nitrogen balance, and HOMA index were similar between the two treatment groups.

Plasma lactate and pyruvate levels increased in controls ($p < 0.01$ for both) but decreased in the supplemented group ($p < 0.01$ and 0.02 respectively). EAA supplemented patients but not controls improved both exercise output and peak oxygen consumption and walking test.

Conclusions:

Adequate EPI when combined with essential amino acid supplementation may improve nutritional and metabolic status in most muscle-depleted CHF patients.

1. Introduction

The prognostic importance of increasing body weight (BW) in patients with chronic heart failure (CHF) has been confirmed by recent studies showing that patients with a body mass index (BMI) of 30-34.9 kg/m² have a lower mortality rate than those with a BMI < 30 kg/m² [1] [2] [3] [4].

Nutrition is obviously the natural way of increasing BW. However, we suspect that even if adequate, nutrition as energy-protein intake (EPI) alone might not be enough to increase BW in malnourished CHF patients, unless it can also limit possible metabolic derangements in the skeletal muscles [5, 6].

We base our hypothesis on clinical evidence that severe muscle protein malnutrition and normal BW may co-exist (39% of clinically stable CHF patients) [7] and on documented evidence in CHF of skeletal muscle energy shortage [8], which is essential for protein synthesis [9]. In addition, we also asked whether simple nutritional substrates such as essential amino acids (EAAs) could enhance the ability of EPI to increase BW, while also attenuating metabolic abnormalities in skeletal muscles. The rationale behind using EAAs derives from the documented ability of amino acids to regulate protein turnover by acting on protein metabolism [10], increasing aerobic metabolism [11] and increasing the biological activities of anabolic hormones [12, 13] such as insulin and insulin-like growth factor-1 (IGF-1).

We therefore wanted to identify certain metabolic aspects of muscle-depleted normal weight patients with CHF and to investigate whether EAA supplements could improve both BW/muscle mass and metabolic derangements in muscles over time.

2. Methods

2.1. Population

Forty-four clinically stable CHF outpatients were enrolled in the study. Patients were selected from two clinical databases according to the following criteria:

1. normal BW (= a body mass index (BMI) $>20 < 25 \text{ kg/m}^2$ which had been stable over the past year) and high depletion of skeletal muscle mass (arm muscle area, AMA; $< 10^\circ$ percentile of normal values for age and sex) [14],
2. stable daily EPI over the past year, providing energy $\geq 30 \text{ kcal/kg}$ and proteins $> 1.1 \text{ g/kg}$ [7]. This was assumed to be adequate as it was associated with a positive nitrogen balance (NB), that is $> +1 \text{ g/24 h}$, indicating a predominance of anabolic processes [7],
3. adequate daily physical activity.

According to our clinical practice, anthropometric variables, nutritional intake, and nitrogen balance were checked every 4 months.

Patients with diabetes mellitus treated with hypoglycaemic drugs or insulin therapy, with liver and renal insufficiency were excluded from the study. Clinical stability of CHF was assessed according to the absence of fluid retention (peripheral or pulmonary), normal jugular venous pressure and no changes in medication over the past three months.

2.2. Study procedures

At 8 am, after a 12-h overnight fast, plasma concentrations of lactate [15], pyruvate [16] ($\mu\text{mol/ml}$) and serum insulin ($\mu\text{U/ml}$) [7] were measured from peripheral venous blood samples in all patients. The plasma lactate/pyruvate ratio (L/P) was also calculated. These specific acids were assessed because elevated levels in the plasma may be caused by alterations in the regulation of the glucose metabolic pathway [17] and not just by hypoxia which was not present in our patients.

Insulin resistance was estimated using the Homeostasis Model Assessment (HOMA) [18] as follows:

$$\text{HOMA} = \frac{\text{blood glucose level (mmol/l)} \times \text{fasting serum insulin level } (\frac{\mu\text{l}}{\text{l}})}{22.5}$$

The normal value of HOMA in healthy elderly patients aged 61 to 81 years is < 2.4 [19].

All patients had their NB calculated as described previously [10]. Urinary nitrogen analysis to calculate NB was made using a urease/glutamate

dehydrogenase coupled enzymatic technique (BUN method, Urea Nitrogen Flex[®] reagent cartridge, Dade Behring).

Nutritional intake was measured using a 7-day food diary [7]. Each patient was asked to keep a food diary at home for 7 consecutive days, before and after meals, to record the type and weight of cooked or uncooked food which they consumed. We converted this data to raw equivalents using appropriate tables when necessary [7,20].

Patients became accustomed to keeping a nutritional diary as an essential part of their clinical control over time. Moreover, we regularly reminded the patients of the importance of keeping an accurate diary for this study. The principal investigator was responsible for keeping the patients informed, for checking the food diaries and for calculating nutritional intakes (using a computer program set up by our group) [21]. Energy (kcal) and macronutrient intake (carbohydrates, proteins, and lipids expressed in g) were also related to the patients' body weight (kg). Energy was also expressed as a percentage of basal metabolic rate (BMR) [21].

After completing the baseline evaluations, the patients underwent a 6-min walking test [22] and bicycle exercise test. All exercise tests were performed in the morning after an overnight fast of between 12-16 h. We adopted a RAMP protocol in which the work load increase was 10 W/min with pedalling at 40 to 50 rpm. The patients were connected to a system for respiratory gas analysis, as described elsewhere [23]. Patients were asked to exercise until exhausted. Among cardio respiratory variables, we measured oxygen peak (VO_2 peak) and respiratory exchange ratio (VCO_2/VO_2).

Patients were then assigned to treatment according to a randomised allocation procedure. A randomisation list was generated using SAS statistical software, A and B being the identifiers of the blinded treatment. The list was made available to both the cardiologists (C.O., A.G., A.T., and E.V.) and the hospital pharmacist. The cardiologists allocated patients sequentially to treatment A or B according to the randomisation list. The first author of the article (A.R.) who interpreted all the results was blinded to the treatment allocation. Supplemented patients were informed by the cardiologists that the amino acid supplement served to implement their dietary intake; the patients did not receive any information about the reason for giving the supplement.

Twenty two patients (EEA group) received an oral nutritional mixture supplement, which provided 8 g of essential amino acids (EAAs) per day; (Aminotrofic, Professional Dietetics, Milan, Italy; see Table 1) (4 g in the morning +4 g in the afternoon diluted in half a glass of water). The remaining 22 patients received no supplementation (control group). The amount of nitrogen in the EAA supplement (1.28 g) was added to the nitrogen intake from the diet, in order to calculate the total nitrogen intake for supplemented patients.

Table 1. Nutritional composition of a single packet containing 4 g of an amino acid mixture

kcal	35.3
kJ	149.9
Total amino acids of which	4 g
L-Leucine	1250 mg
L-Lysine	650 mg
L-Isoleucine	625 mg
L-Valine	625 mg
L-Threonine	350 mg
L-Cysteine	150 mg
L-Histidine	150 mg
L-Phenylalanine	100 mg
L-Methionine	50 mg
L-Tyrosine	30 mg
L-Tryptophan	20 mg

After 2 months, patients underwent the same clinical, functional biochemical and nutritional (7-day food diary) investigations as at baseline. Moreover, BW was recorded and AMA was calculated [14]. Patients were weighed with underwear only, between 9 am and 10 am. The baseline and 2 month anthropometric measures were both assessed by the same investigator. The coefficient of variability of triceps skin fold thickness and mid-arm muscle circumference, necessary to calculate AMA, was 1%.

Given that the normal daily variation of BW is 0.5 ± 0.2 kg [24], we considered an increase in body weight greater than 1 kg ($BW > 1$ kg) over 2-months, as significant. We assumed that an increase in skeletal muscle mass (AMA) was significant over 2 months when it was associated with a BW increase of > 1 kg.

This study was approved by the Technical Ethics Committee of Foundation “S. Maugeri”, Pavia, Italy and written informed consent was obtained from all participants after the nature of the study had been explained.

2.3. Evaluation of patient compliance with EAAs

Patient compliance was estimated by counting the number of amino acid packets remaining 2 months after the start of the protocol (each patient was given 60 packets of EAA supplement). This allowed us to estimate patient compliance over the 2-month study period. In order to judge patient reliability (at 2-months), we also took venous blood samples for measurement of fasting plasma leucine concentration (the main amino acid in the packet). Importantly, the blood samples were drawn 60 min after the patient should have taken the morning packet of EAAs. The patients were not informed of their plasma leucine measurement.

Patient reliability was judged based on plasma leucine concentrations. In our laboratory, for healthy elderly subjects, the increase in baseline plasma leucine 60 min after 4 g EAA ingestion was +145%. The same procedure was also applied to the control patients.

2.4. Statistical analysis

The χ^2 test was used to compare the distribution of demographic, clinical, functional and treatment characteristics as well as of a BW increase >1 kg between the two patient groups. Comparisons between groups at baseline and after two months from the start of the protocol were performed using ANOVA. Results are shown as a mean value \pm SD. Differences were considered statistically significant at $p < 0.05$.

3. Results

Fig. 1 is a flow chart of patient participation in the EAA supplementation study. There were five dropouts in the control group and one in the intervention groups. Thus, the analysis included 38 patients (17 controls +21 supplemented). The demographic, clinical functional and treatment characteristics of the patients at baseline are summarised in Table 2.

Table 2. Demographic, clinical, functional and treatment characteristics at baseline

	PATIENTS $n = 38$		
	Controls	EAAs	p value
Number (n)	17	21	ns
Age (yr)	73.1 \pm 2.8	74.5 \pm 5	ns
Men/women	14/3	13/8	ns

	PATIENTS <i>n</i> =38		
	Controls	EAA's	<i>p</i> value
Aetiology			
Coronary artery disease	7 (41.2%)	12 (57.1%)	ns
Valvular heart disease	2 (11.7%)	3 (14.3%)	ns
Idiopathic	8 (47%)	6 (28.6%)	ns
Duration of disease (months)			
<18	7	4	ns
≥18	10	17	ns
NYHA class			
I	0	0	ns
II	12 (70.6%)	15 (71.4%)	ns
III	5 (29.4%)	6 (28.6%)	ns
IV	0	0	ns
Left ventricular ejection fraction	29.7±9.2%	29.4±17.4%	ns
Medication			
ACE inhibition	17pts (88.2%)	21pts (100%)	ns
β blockers	15pts (88.2%)	20pts (95.2%)	ns
Digoxin	7pts (41%)	8pts (38%)	ns
Diuretics	17pts (100%)	21pts (100%)	ns



Figure 1. Flow chart of participation in the nutritional supplementation study.

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Table 3 shows the nutritional intake of patients in both treatment groups.

Table 4 shows the nutritional status, nitrogen balance, plasma levels of glucose, lactate, and pyruvate, serum insulin concentrations, and insulin resistance from HOMA in both patient groups at baseline and at 2-months.

Table 3. Nutritional intake in the two groups of patients at baseline and after 2 months

	Controls		EAA supplemented	
	Baseline	2-mo	Baseline	2-mo
Energy				
kcal	1884±205	1950±279	1890±250	1950±300
kcal/kg	31±2.9	32.3±3.2	33.7±3	33.5±2.7
% BMR	132%	-	136%	-
Carbohydrates				
g	222±36	239±21.5	263±90	303±115
g/kg	3.65±0.6	3.9±0.9	4.7±1.8	5.2±2.1
Proteins				
g	68±9.6	67±9.1	78±11	76±27
g/kg	1.12±0.16	1.1±0.15	1.4±0.2	1.3±0.5
Lipids				
g	73±12	79.6±24	64.3±27	76±40
g/kg	1.2±0.2	1.3±0.4	1.15±0.5	1.3±0.7
Fatty acids (% energy)				
Saturated	11.5±3	13±4	12±2	11.8±4

a Data are expressed as mean±standard deviation (SD).

b BMR = basal metabolic rate.

c Statistical analysis (Anova test and Fisher's PLSD test): No differences in macro-micronutrients were found between the two subgroups and respectively within each subgroup neither at baseline nor at 2 months.

a Water content in food.

	Controls		EAA supplemented	
	Baseline	2-mo	Baseline	2-mo
Monounsaturated	18.5±2.5	16±3.9	14±3	13±5
Polyunsaturated	5±2	7.7±3	4.5±2	10.3±3
Alcohol (g)	19±5	17±7	16±4	20±6
Fibre (g)	15.4±5	17±8	22±9	20±5
Calcium (mg)	1001±350	17±8	22±9	20±5
Phosphorus (mg)	1226±420	1021±500	850±324	1456±645
Potassium (mg)	2590±185	2894±350	2330±155	2950±284
Sodium (mg)	1401±155	1404±107	1258±270	1200±159
Iron (mg)	10.5±4	9.2±3	12±5	11.5±4
Zinc (mg)	10.2±3	9.5±4	9±4	9.8±3.5
Thiamine (mg)	0.5±0.1	1.2±0.3	1.02±0.15	1.07±0.22
Riboflavin (mg)	1.3±0.2	1.1±0.3	1.15±0.2	1.35±0.4
Niacin (mg)	15.5±4.8	17.7±5.2	17.9±3.2	16±4.1
Vit A (µg)	650±135	705±250	594±101	674±189
Vit C (mg)	66±32	75±28	79±15	85±25
Tocopherol (mg)	6.5±3	8.2±4	7.1±3.5	7.9±4.1
Colecalciferol (µg)	1.95±0.8	2.1±0.9	2.1±1	2.4±1.1
Water (ml) ^a	1113±155	990±250	1005±175	1120±148

a Data are expressed as mean±standard deviation (SD).

b BMR = basal metabolic rate.

c Statistical analysis (Anova test and Fisher's PLSD test): No differences in macro-micronutrients were found between the two subgroups and respectively within each subgroup neither at baseline nor at 2 months.

a Water content in food.

Table 4. Nutritional status, nitrogen balance, insulin and glucose concentrations, insulin resistance (HOMA index), plasma lactate and pyruvate concentrations, lactate/pyruvate ratio in two subgroups of patients at baseline and 2 months after

	Controls		EAA supplemented	
	Baseline	2-mo	Baseline	2-mo
Body weight kg	60.8±7	61.2±6.3	55.9±17	58.2±7.2 [^]
Body mass index kg/m ²	23.2±1.4	23.6±1.5	22.5±2.1	23.4±1.9 [^]
Triceps skin fold thickness mm	11.9±3.7 (6) ^a	11.4±3.7	10.4±4.4	10.3±3.9
Arm muscle mass area cm ²	34.2±5 (44.7) ^a	37.1±4 ^{^^}	31.2±9.9	34.9±10 ^{^^}
Nitrogen balance g/24 h	3±2.8	2.8±2.5	3.9±3.3	3.6±3.3
Plasma glucose mg/dl	90±13	96±8	95±26	97±21
Serum insulin μU/ml	21.4±6.6 (nv 10±3.8)	21.3±6.7	18±15	16±12
HOMA index	4.77±1.8 (nv 2.1±0.2)	5.08±2	4.31±3.2	3.7±2.5 ^{^^^}
Plasma glyconeogenic substrates				
Lactate (μmol/ml)	1.1±0.4 (nv 0.2±0.15)	1.27±0.4 [^]	1.2±0.3	0.9±0.35 ^{^°°}

a Data are expressed as mean±standard deviation (SD).

b nv: normal value of laboratory.

c Statistical analysis (Anova test and Fisher's PLSD test):

d Intra groups differences (2 mo vs baseline):

[^] $p < 0.01$

^{^^} $p < 0.02$

^{^^^} $p < 0.06$.

h Control group 2 mo vs EAA supplemented group 2 mo:

^{°°} $p < 0.001$.

a Values corresponding to 10^o percentile.

	Controls		EAA supplemented	
	Baseline	2-mo	Baseline	2-mo
Pyruvate (µmol/ml)	0.018±0.009 (nv 0.015±0.008)	0.027±0.012 [^]	0.02±0.01	0.015±0.008 ^{^^ °°}
Plasma lactate/pyruvate ratio	61±44 (nv 14±10)	47±31	60±30	60±25

a Data are expressed as mean±standard deviation (SD).
b nv: normal value of laboratory.
c Statistical analysis (Anova test and Fisher's PLSD test):
d Intra groups differences (2 mo vs baseline):
[^] $p < 0.01$
^{^^} $p < 0.02$
^{^^^} $p < 0.06$.
h Control group 2 mo vs EAA supplemented group 2 mo:
^{°°} $p < 0.001$.
a Values corresponding to 10[°] percentile.

3.1. Baseline characteristics of the study population

According to our selection criteria, the two groups of patients had similar body mass index (BMI) and reduction in skeletal mass (AMA). Subcutaneous fat tissue (TST) was within normal values for age and sex and was similar for both groups. Moreover, daily nutritional intakes were not significantly different between the two groups. Both groups had similar positive nitrogen balance (NB=>+1 g/day) indicating that the patients' body metabolism was oriented towards the anabolic state.

Both groups had a similar state of insulin resistance (HOMA>2.4). Plasma lactate and pyruvate concentrations as well as L/P ratio were far higher in both groups than previously determined in our laboratory for healthy elderly subjects. There were no differences in respiratory variables or physical capacities between the two groups (Table 5).

Table 5. Respiratory variables both at rest and during exercise (bicycle) and 6-minute walking test results for the study patients at baseline and after 2 months (mean±SD)

	Controls		EAAs supplemented	
	Baseline	2-mo	Baseline	2-mo

	Controls		EAAs supplemented	
	Baseline	2-mo	Baseline	2-mo
A) Cycle ergometer testing				
Power output (W)	85±24	88±22	80±28	95±25 ^{** ^}
Respiratory variables				
1) VO ₂ (ml O ₂ /kg/min)				
Rest	4.2±0.6	4.4±0.8	4.5±0.8	4.2±1
Peak	12.9±2.7	13±3.5	13.5±1.7	14.9±1.9 ^{* ^^}
2) Respiratory exchange ratio				
Rest	0.88±0.01	0.95±0.1	0.87±0.08	0.96±0.06
Peak	1.1±0.09	1.12±0.1	1.1±0.2	1.15±0.2
B) 6-min walking test (m)	298±142	310±155	331±124	405±130 ^{*** ^}
a Inter group differences in time courses of:				
* $p < 0.05$ oxygen consumption.				
** $p < 0.01$ work output.				
*** $p < 0.001$ walking ability.				
e Intra group differences (2 mo vs baseline):				
^ $p < 0.02$				
^^ $p < 0.05$.				

3.2. Patient compliance to EAA supplementation

The results showed that the intervention group very probably complied with the EAA supplementation. Indeed, none of the patients returned any packets at the end of the 2 month study period. In addition, the increase in plasma leucine concentration at 2 months was 279% of the baseline value ($128 \pm 28 \mu\text{mol/L}$ at baseline vs $478 \pm 104 \mu\text{mol/L}$ at 2 months, $p < 0.001$), which suggests patient reliability. In the control patients, no significant changes were observed in plasma leucine concentration between baseline and 2-months ($115 \pm 22 \mu\text{mol/L}$ at baseline to $121 \pm 31 \mu\text{mol/L}$).

3.3. Patient evaluation after two months of study treatment

No significant changes in New York Heart Association (NYHA) functional

class, clinical, nutritional intake, or functional characteristics were found between the two groups (data not shown).

A BW increase of more than 1 kg was found in 80% of the supplemented group (from 58 ± 4 kg to 61 ± 3 kg; $+2.96 \pm 1.56$ kg, $p < 0.01$ compared with baseline) and in 30% of the non-supplemented group (from 59.9 ± 3.5 kg to 62.2 ± 3.2 kg; $+2.3 \pm 0.8$ kg, $p < 0.05$ vs baseline). This difference in distribution between the two treatment groups was significant ($p < 0.05$). Since there was a different male:female ratio between the two treatment groups, we calculated the sex related differences in BW improvements within each group and found that they were similar. Indeed, the BW gain in the EAA group was $+3.3 \pm 2.4$ kg in males and $+2.7 \pm 0.6$ in females (ns). In the control group, males gained $+2 \pm 0.7$ kg and females $+2.6 \pm 0.9$ kg (ns). No patient, regardless of treatment group, had a reduced BW at baseline. Baseline skeletal muscle mass (AMA) increased significantly and similarly in both groups. No patient, regardless of group had a reduced AMA over time. Baseline subcutaneous fat tissue (TST) in both groups remained virtually unchanged.

Insulin resistance (HOMA) tended to deteriorate in the non-supplemented CHF patients (from 4.7 ± 1.8 to 5.1 ± 2 ; $+15\%$; $p = 0.06$) while it improved in the EAA group (from 4 ± 3.2 to 3.7 ± 2.6 -15% ; $p = 0.07$). However, this difference was not significant.

The effect on plasma lactate and pyruvate was different in the two treatment groups. Indeed, lactate increased in all control patients (from 1.1 ± 0.4 $\mu\text{mol/ml}$ to 1.27 ± 0.4 $\mu\text{mol/ml}$; $p < 0.01$) but decreased in all supplemented ones (from 1.2 ± 0.3 $\mu\text{mol/ml}$ to 0.9 ± 0.35 $\mu\text{mol/ml}$; $p < 0.01$). The difference in effect between the two groups was highly significant ($p < 0.001$).

Like lactate, plasma pyruvate concentration increased in all control CHF patients (from 0.0018 ± 0.009 to 0.027 ± 0.012 $\mu\text{mol/ml}$; $p < 0.01$) but decreased in all EEA CHF patients (from 0.02 ± 0.01 to 0.015 ± 0.008 $\mu\text{mol/ml}$; $p < 0.02$). Plasma L/P ratio tended to decrease in the controls but remained unchanged in the EAA group.

After 2 months, patients in the EAA group significantly improved their bicycle exercise capacity ($+15 \pm 7$ W, $p < 0.02$ vs basal value), peak oxygen consumption (VO_2 peak $+1.1 \pm 0.3$ ml/kg/min, $p < 0.05$ vs basal value) and walking capacity ($+74 \pm 25$ m, $p < 0.02$ vs basal value) (Table 5). There was no improvement in the control group.

In summary, the results of this study show that adequate EPI alone was associated over time with: a) a BW increase of more than 1 kg in 30% of CHF patients; b) a tendency to worsening insulin resistance; c) increased plasma lactate and pyruvate levels; and d) unchanged exercise and walking capacities. The combination of adequate EPI + 8 g EEAs was associated with a) a BW increase of more than 1 kg in 80% of CHF patients; b) a trend towards an improvement in insulin resistance; c) decreased plasma lactate

and pyruvate levels; and d) improved exercise and walking capacities.

4. Discussion

This study shows that in muscle-depleted, normal weight, clinically stable patients with CHF, alterations in muscle metabolism may co-exist with an anabolic body state. The results show that adequate EPI increases BW by more than 1 kg in less than one third of patients and moreover, it may not limit metabolic disorders in the muscle, but may indeed exacerbate the problem over time. Conversely, if enriched with EAAs, adequate EPI can increase BW by more than 1 kg in up to 80% of patients and improve metabolic disorders in the muscle.

4.1. Baseline muscle metabolic disorders and anabolic state

This study shows that in clinically stable muscle-depleted CHF, an adequate EPI was associated with an anabolic state, but did not correct alterations in muscle glucose oxidation.

Elevated plasma lactate and pyruvate levels suggest reduced muscle capacity to oxidize pyruvate (glucose) rather than an excess of glucose anaerobic breakdown due to the rate of oxygen use [25]. A number of conditions associated with CHF including hormone imbalance [7,26], insulin resistance [27 [28] 29] and cytokine overproduction [25,28] can reduce pyruvate oxidation by inactivating the pyruvate dehydrogenase complex (PDH), the rate-limiting step for lactate and pyruvate entry into the citric acid cycle for oxidation [30,31].

In particular, cytokine overproduction may play a major role in perturbing muscle aerobic glucose metabolism. For instance, chronic intravenous infusion of interleukin 1 (IL-1) has been found to cause a 40% reduction in active form-PDH in the skeletal muscle of healthy rats [31]. This metabolic effect has been shown to induce elevated plasma lactate concentrations similar to those observed in patients and animals with sepsis [32 [33] [34] 35].

Moreover, modulation of the tumour necrosis factor response abrogates muscle lactate production and hyperlactatemia during hypermetabolic sepsis [36,37].

The reduced pyruvate oxidation leading to accumulation of glycolytic intermediates, cellular acidosis and reduced energy reserves, are all metabolic disturbances that can potentially have an impact on nutritional status. Indeed, cellular acidosis can induce both somatic and myofibrillar proteolysis [38] and cell energy shortage may induce reduced protein synthesis [9].

The mechanisms leading to increased lactate formation and reduced pyruvate oxidation could also explain the co-existence of muscle depletion

and fat tissue/body weight conservation observed in our study patients. For instance, at an adipose tissue level the lipolytic activity of high cortisol levels [7,39] can be counteracted by increased serum insulin concentration, however in muscles the proteolytic effect of cortisol contributes to cellular metabolic derangements. Again, while the conservation of fat tissue may be favoured by beta-blockers [2,3], a reduction of protein synthesis might also be due to growth hormone resistance [26] and insulin-like growth factor-1 (IGF-1) reduction [40]. As the patients in our study had optimal nutritional intake and clinical conditions, it is conceivable that any disturbances in their current anabolic state and/or a further worsening disorders of muscle metabolism, for instance from large reductions in nutritional intake and/or metabolic stress (infection, haemodynamic instability) may bring about a global loss of body weight [41].

4.2. Two months after randomisation

This study clearly shows the nutritional and metabolic benefits for patients of EAAs supplements and suggests that the improvement in nutritional status may be a consequence of the well documented metabolic activities exerted by EAAs. Indeed, EAAs can influence both protein turnover and aerobic metabolism in muscle (Fig. 2).



Figure 2. Some documented actions of amino acids potentially useful to improve both nutritional and metabolic status of patients with CHF.

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Concerning protein turnover, EAAs act on protein metabolism by both direct and indirect mechanisms. Directly, EAAs stimulate protein synthesis [42] and inhibit proteolysis [43] in several tissues including skeletal muscle, heart, liver, and pancreatic β -cells. It is worth noting that EAAs can promote protein synthesis independently of insulin [10] which is relevant for insulin resistant CHF. Indirectly, EAAs may stimulate protein synthesis by increasing biological activity of insulin-like growth factor 1 (IGF-1). Levels of IGF-1, which have been shown to be low in both cachectic [26] and non-cachectic [40] CHF patients, are positively associated with muscle protein synthesis [44] and negatively associated with protein breakdown [45].

The stimulation of aerobic metabolism in muscle [9, 11] (Fig. 2) is the other mechanism by which EAAs can improve both metabolic derangements and at the same time nutritional status. Here again, EAAs may act by both direct

and indirect mechanisms.

EAA directly influence the aerobic pathway, as they can enter it at various levels and can be used as an alternative fuel for energy production [9]. The reactivation of aerobic pathway activity makes the lactate \rightleftharpoons pyruvate reaction shift to the right [9]. In this way, lactate formation diminishes and more pyruvate is processed in the aerobic pathway. This may explain the reduction in plasma lactate and pyruvate levels observed in CHF patients after 2-months of EAA supplementation.

EAA could improve glucose oxidation indirectly by reducing insulin resistance [12,46]. Thus, EAA can re-modulate the muscle cell glucose breakdown/use ratio. The improvement in both nutritional state and metabolic abnormality may explain increased exercise output and oxidative metabolism as well as walking capacity in supplemented patients. The increased respiratory capacity and walking test confirm the findings of both a previous [47] and a very recent investigation [23] that used the same substrates (EAA), at the same dose (8 g/d) in a similar elderly population.

It is conceivable that in our study, EAA may have favoured patient tissue accrual by harmonizing protein synthesis processes and cell energy production essential for protein synthesis. In this way, EAA seems to act as a bridge between nutritional intake and muscle metabolism.

The fact that an adequate EPI alone cannot reduce metabolic disturbances in muscle, suggests that amino acids from food proteins are not sufficient and/or capable of attenuating these metabolic muscle defects. This might limit protein synthesis and tissue accrual in most non-supplemented patients.

In contrast, since adequate EPI alone is associated over time with an increase in muscle disturbances we suspect that attempts to excessively force nutrition in CHF in order to increase body weight could induce/increase insulin resistance and worsen the metabolic state of patients.

5. Clinical implications

In light of a very recent investigation showing the importance of maintaining BMI between 30 and 34.9 kg/m² in order to reduce mortality in patients with CHF [4], our results, particularly if corroborated by a larger clinical trial, may offer one way of improving BMI. According to a recent report by Anker et al., our patients could be defined as “sarcopenics” [41]. In sarcopenic patients, the combination of adequate nutrition and EAA might prevent the transition from a sarcopenic condition to a global loss of body weight [41].

The determination of plasma lactate and pyruvate levels both at baseline and over time offers the physician a rapid understanding of both muscle metabolism and its time course after pharmacological and nutritional

interventions. Increased plasma lactate and pyruvate concentrations should not be related to cellular hypoxia alone, but may also be considered as products of altered cell substrate use. In this case, an otherwise adequate EPI should not be increased but rather amino acids supplementation should be considered.

6. Limitations

For technical reasons we did not use dual X-ray absorptiometry (DXA) methods to measure body composition. This would have strengthened our discussion. Another method we could have used to assess body composition is bioelectrical impedance analysis (BIA) and more importantly, bioimpedance spectroscopy (BIS) which provides an estimate of body cell mass. However, we preferred not to use these methods because they can be problematic for individual assessment in the clinic, particularly in CHF patients with abnormal fluid distribution or body geometry [48].

The male:female ratio was not equally represented between the two groups of patients. However this did not limit the interpretation of the data as the weight changes in patients achieving a >1 kg increase in BW at 2-months were similar for males and females in each group. Moreover there was a similar BW gain when males and females were compared between the two groups.

At first glance, the selection of patients based on their specific nutritional intakes (energy ≥ 30 kcal/kg+proteins > 1.1 g/kg) might seem to be a limitation of this study. However, we believe that this is a strength, since choosing patients with stable nutritional intake over time allows a better balance between nutrition and metabolism and hence, a more reliable link between them.

Plasma lactate and pyruvate levels might not automatically reflect their relative tissue concentrations. However, in the absence of peripheral oedema, particularly in the arms, from where the blood samples were taken, it is reasonable to assume that the passage of EAAs from the cells to the blood stream occurred without any difficulty.

Finally, plasma cytokine levels were not determined. This is a major issue as a recent study showed that inflammatory cytokines, together with weight loss and reduced food intake, are involved in inducing cachexia and poor functional status and prognosis [49].

7. Conclusions

This study shows that it is possible to improve nutritional status and muscle metabolic derangements in muscle-depleted normal weight patients with CHF, by combining an adequate energy-protein intake with essential amino acid supplementation.

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